

Role of Gibberellic Acid in Abolishing the Detrimental Effects of Cd and Pb on Broad Bean and Lupin Plants

Abd El-Monem M. Sharaf, Ibrahim I. Farghal and Mahmoud R. Sofy

Botany & Microbiology Department, Faculty of Science, Al-Azhar University, Cairo, Egypt.

Abstract: This work describes the changes in photosynthetic pigments, soluble carbohydrates, proteins and activities of certain enzymes of both broad bean and lupin plants in response to heavy metals (Cd or Pb), gibberellic acid (GA₃) and their interactions. Significant reduction in contents of chlorophylls, soluble carbohydrates, soluble proteins were recorded in tested plants due to the treatment with either Cd or Pb. Application of GA₃, in most cases, decreased contents of chlorophylls and soluble carbohydrates in absence and presence of either Cd or Pb, while contents of soluble proteins were markedly increased. All applied treatments greatly affected the activities of amylases, proteases, catalases and peroxidases in both broad bean and lupin plants. Generally, it could be concluded that GA₃ have (to some extent) a beneficial regulatory role in plants grown under both Cd and Pb polluted conditions.

Key words: Broad bean, lupin, gibberellic acid, lead, cadmium.

INTRODUCTION

Cadmium and lead are emitted from metal working and manufacturing industries and from cars. The problem in Egypt is that most industrial areas are located close to the cultivated lands. Moreover, roads and high ways carry a vast number of motor vehicles running through the cultivated areas in Egypt. Consequently, these heavy metals, once taken up by plants, enter human food chains^[19,22]. The uptake of trace elements from aerial sources through the leaves may have a significant impact on plant contamination especially of elements such as Cd and Pb^[31].

Metal tolerance is a phenomenon often noted, but the understanding of the physiological and biochemical aspects of these fascinating aspects of plant metabolism has not progressed a pace. The use of growth regulators is promising for enhancing and improving the production of economic food crops. Recently, these substances could not only regulate plant growth and development but also increase plant resistance to various environmental stress conditions^[14,12]. The present study was undertaken to investigate the effects of heavy metals (Cd or Pb), GA₃ and their interactions on certain metabolic activities of broad bean and lupin plants.

MATERIALS AND METHODS

Seeds of broad bean "*Vicia faba*" (Giza-716) & lupin "*Lupinus albus*" (Giza-1) were obtained from Agricultural Research Centre, Ministry of Agriculture, Giza, Egypt. Uniform broad bean and lupin seeds were

planted in Botanical garden, Botany and Microbiology Dept., Fac. of Sci., Al-Azhar Univ., in two plots containing 12 ridges (6 for each plot). The seeds were sown on one side of the ridge, with 20 cm apart between the hills. The developed plants were irrigated when ever required. Each of the two experimented plants was divided into 6 groups representing the foliar treatments. water (control), 20 ppm Cd as cadmium chloride, 100 ppm Pb as lead acetate, 100 ppm GA₃, 20 ppm Cd + 100 ppm GA₃ and 100 ppm Pb + 100 ppm GA₃. The plants were treated twice with the above mentioned treatments. In cases of the combined treatment, GA₃ was sprayed firstly followed by heavy metal with 24 h interval. The first treatment was made when the age of plants was 33 days; while the second treatment was made the age of plants was 65 days. The plant samples were collected for analysis when the plants were 72 days old. At the end of the growth season, analysis of the seeds yielded from the different treatments and the control were measured. Contents of chlorophylls were estimated using the method of Vernon and Selly^[33]. Contents of soluble carbohydrates were measured according to the method of Umbriet *et al.*^[32]. Contents of soluble proteins were estimated according to the methods of Lowry *et al.*^[20]. Activities of amylases, were determined using the method of Afifi *et al.*^[3]. Proteases activities were estimated using the method of Ong and Guacher^[24]. Catalases activity was measured according to the method of Chen *et al.*^[7]. Peroxidase activity was assayed according to the method of Bergmeyer^[5]. Statistical analysis of the obtained results was done using (L.S.D.) according to Snedecor and Cochran^[30].

RESULTS AND DISCUSSION

Photosynthetic Pigments: Results presented in Table (1) showed that, in broad bean plants, contents of chlorophyll (a) as well as total chlorophyll (a + b) were, mostly, decreased due to the application of GA₃, however, contents of chlorophyll (b) were significantly increased. In lupin plants, treatment with GA₃ caused, generally, significant increases in the contents of chlorophyll (a), (b) and total chlorophyll (a + b) contents.

Increases in photosynthetic pigments that observed in the present work in response to GA₃ agreed to those observed by El-Khateeb ^[10] working on broad bean and soybean, Abd El-Hamid *et al.* ^[1] working on sugar beet, Abdel-Hameid ^[2] working on safflower plants and Picazo *et al.* ^[25] working on rice plants. The present study showed that, contents of chlorophyll (a), (b) and their contents in both broad bean and lupin plants were significantly reduced in response to the application of either Cd or Pb. Similar results were reported by several other investigators ^[23,11,1,21,4,29].

With respect to the combined treatments of GA₃ with either Cd or Pb, the obtained results showed different responses as regards the contents of chlorophylls of the two tested plants. In broad bean plants, application of GA₃ was found to be, mostly, ineffective in overcoming the adverse effects of both Cd or Pb regarding the contents of chlorophyll (a), (b) and total (a + b) chlorophyll contents. However, in lupin plants treated with either Cd or Pb, application of GA₃ caused, generally, slightly increment in the chlorophyll (a), (b) and total chlorophyll contents. These results are in accordance with other investigators; Abd El-Hamid *et al.* ^[1] found that, in sugar beet plants, treatment with GA₃ declined the adverse effects of either Cd or Pb as regards the level of different photosynthetic pigments. Also, Picazo *et al.* ^[25] found that, the net photosynthesis rates recovered transitorily in Cd-treated rice plants after the addition of GA₃ (14 µm).

Carbohydrates: Table (2) showed that, contents of soluble carbohydrates in shoots as well as in the yielded seeds of broad bean plants were, highly significantly decreased in response to the application of GA₃. However, opposite trends were recorded in lupin plants, where as contents of soluble carbohydrates in shoots and also, in the yielded seeds were markedly increased in response to the treatment with GA₃. In this regard Abdel-Hameid ^[2] reported that, spraying safflower plants with GA₃ caused highly significant increases in the contents of total soluble carbohydrates

in all parts (leaves, stems, roots and yielded seeds). Also, BiaECKa and Kepezynski ^[6] found that, application of GA₃ increased the contents of total soluble carbohydrates of *Amaranthus caudatus* plants.

In the present study, it was found that, contents of total soluble carbohydrates in shoots, roots and yielded seeds of both broad bean and lupin plants were highly significantly decreased in response to the application of Cd or Pb. These results are in agreement with those obtained by several other investigators ^[28,11,1,8,21,4,25,27].

With respect to the combined treatments, results of the present work (Table 2) revealed that, in both broad bean and lupin plants, application of GA₃ with either Cd or Pb resulted in, mostly, significant decreases as regards the contents of total soluble carbohydrates in shoots, roots as well as in the yielded seeds. These indicate that, application of GA₃ was not effective in overcoming the adverse effects of Cd and Pb as regards the contents of total soluble carbohydrates of broad bean and lupin plants. On the contrary, the obtained results, Abd El-Hamid *et al.* ^[1] reported that, the use of growth regulators, especially GA₃ seemed to have pronounced effect on the level of sugars fractions of sugar beet plants and could be used for minimizing harmful effects of Cd or Pb. Also, Picazo *et al.* ^[25] found that, GA₃ increased the sugar content in roots, second and third leaves and also, modified the carbohydrate distribution pattern of rice plants grown under the application of Cd or Ni.

Protein contents: Results of the present work, Table (3) revealed that, in both broad bean and lupin plants, treatment with GA₃, in absence and presence of either Cd or Pb, resulted in, mostly, highly significant increases in protein contents in shoots, roots and also, in the yielded seeds. These results are in harmony with those obtained by Khalaf *et al.* ^[17] on soybean, common bean and cowpea plants where they found that, contents of proteins increased as a result of GA₃ application. Abdel-Hameid ^[2] reported that, the contents of soluble proteins of the different parts of safflower plants were, mostly, highly significantly increased in response to the treatment with GA₃. On the contrary, Gaber *et al.* ^[13] found that, application of GA₃ decreased the contents of soluble proteins in the yielded seeds of *Vicia faba* plants.

On the other hand, the obtained results showed that, application of Cd or Pb caused significant decreases as regards the contents of soluble proteins in shoots, roots as well as in the yielded seeds of both broad bean and lupin plants. These results are in a coincident with several others ^[16,8,21,27].

Table 1: Effect of Cadmium, Lead and GA₃ and their interaction on the chlorophyll contents (mg/g fresh weight) of broad bean and lupin plants .Each value is mean of 3 replicates ± standard error of means.

Broad bean plant			
Treatment	Chlorophyll (a)	Chlorophyll (b)	Total (a+b)
control	6.34 ± 0.05	3.95 ± 0.02	10.89 ± 0.04
Cd	5.43 ± 0.02 **	3.14 ± 0.07 **	8.57 ± 0.09 **
Pb	5.01 ± 0.02 **	2.92 ± 0.02 **	7.93 ± 0.11 **
GA ₃	5.46 ± 0.01 **	4.51 ± 0.12 *	9.97 ± 0.01 **
Cd + GA ₃	5.26 ± 0.04 **	3.32 ± 0.02 **	8.58 ± 0.07 **
Pb+ GA ₃	4.44 ± 0.04 **	2.91 ± 0.01 **	7.34 ± 0.05 **
Lupin plants			
Treatments	Chlorophyll (a)	Chlorophyll (b)	Total (a + b)
Control	8.76 ± 0.15	3.59 ± 0.30	12.35 ± 0.15
Cd	6.49 ± 0.17 **	3.11 ± 0.02	9.60 ± 0.12 **
Pb	7.15 ± 0.03 **	3.91 ± 0.11	11.0 ± 0.05 **
GA ₃	9.90 ± 0.13 *	4.67 ± 0.06 *	14.57 ± 0.07 **
Cd + GA ₃	8.07 ± 0.05 *	4.03 ± 0.10	12.10 ± 0.07
Pb + GA ₃	9.17 ± 0.07 **	4.12 ± 0.02	13.29 ± 0.17**

** *Significant at 5% confidence level; ** Significant at 1% confidence level.

Table 2: Effect of Cd, Pb, GA₃ and their interactions on the total soluble carbohydrates contents (mg/g dry weight) of broad bean and lupin plants. Each value is the mean of 3 replicates ± standard error of means.

Broad bean plants			
Treatment	shoots	Roots	Seeds
control	64.13 ± 0.37	51.21 ± 0.82	80.36 ± 0.27
Cd	49.36 ± 0.11 **	40.36 ± 0.91 **	59.52 ± 0.36 **
Pb	54.21 ± 0.19 **	41.09 ± 0.31 **	55.36 ± 0.18 **
GA ₃	61.20 ± 0.19 **	59.11 ± 0.17 **	68.45 ± 0.19 **
Cd + GA ₃	55.18 ± 0.23 **	47.38 ± 0.09 *	61.24 ± 0.18 **
Pb+ GA ₃	60.11 ± 1.01 *	50.31 ± 0.14	63.27 ± 0.19 **
Lupin plants			
Treatment	Shoots	Roots	Seeds
control	74.24 ± 0.72	54.01 ± 0.37	79.36 ± 0.13
Cd	60.73 ± 0.31 **	40.07 ± 0.28 **	51.28 ± 0.34 **
Pb	59.30 ± 0.57 **	49.29 ± 0.18 **	55.34 ± 0.41 **
GA ₃	79.01 ± 0.07 **	51.26 ± 0.29 *	91.21 ± 0.27 **
Cd + GA ₃	63.52 ± 0.67 **	55.28 ± 0.37	60.78 ± 0.08 **
Pb+ GA ₃	61.36 ± 0.27 **	50.84 ± 0.39 *	59.14 ± 0.24**

***Significant at 5% confidence level; ** Significant at 1% confidence level.

Table 3: Effect of Cd, Pb, GA₃ and their interaction on the total-soluble protein contents (mg/g dry weight) of broad bean plants. Each value is mean of 3 replicates ± standard error of means.

Broad bean plants			
Treatment	Shoots	Roots	Seed
control	101.12 ± 0.28	70.26 ± 0.22	126.21 ± 0.20
Cd	70.11 ± 0.26 **	57.53 ± 0.27 **	89.71 ± 0.17 **
Pb	62.07 ± 0.75 **	63.11 ± 0.94 **	93.76 ± 0.12 **
GA ₃	120.26 ± 0.19 **	83.35 ± 0.07 **	145.26 ± 0.16 **
Cd + GA ₃	118.29 ± 0.24 **	76.08 ± 0.55 **	136.32 ± 0.28 **
Pb+ GA ₃	86.18 ± 0.91 **	79.28 ± 1.02 **	140.19 ± 0.23 **
Lupin plants			
Treatment	Shoots	Roots	Seeds
control	86.11 ± 0.29	72.21 ± 0.07	138.21 ± 0.21
Cd	61.31 ± 0.62 **	60.11 ± 0.56 **	81.37 ± 0.12 **
Pb	60.71 ± 0.15 **	57.04 ± 0.28 **	64.34 ± 0.17 **
GA ₃	110.91 ± 0.94 **	77.02 ± 0.18 **	153.32 ± 0.17 **
Cd + GA ₃	90.43 ± 1.01 *	70.06 ± 0.17 **	150.25 ± 0.16 **
Pb+ GA ₃	96.19 ± 0.06 **	68.21 ± 0.58 **	157.55 ± 0.21 **

**Significant at 5% confidence level ; ** Significant at 1% confidence level.

Enzymes Activities: Data presented in Table (4) revealed that, in broad bean plants, treatment with GA₃ caused increases in the activity of catalase enzymes, while activities of amylase, protease and peroxidase were insignificantly affected. However, in lupin plants activities of amylases, catalases and peroxidases were increased while, activities of proteases were insignificantly affected. The stimulatory effects of GA₃ as regards the activities of some enzymes were recorded by other investigators, Abdel-Hameid^[2] found that, amylolytic and proteolytic activities were significantly stimulated in safflower plants in response to the treatment with GA₃. Also, El-Fouly *et al.*^[9] found that, GA₃ (25, 50 and 100 ppm) increased amylase activity in broad bean plants.

Different responses, in the present work, were recorded as regards the studied enzymes activities of broad bean and lupin plants in response to the application of either Cd or Pb. In broad beans, treatment with Cd caused significant increases in the activities of amylases, catalases and peroxidases, while activities of proteases were insignificantly affected. Treatment with Pb significantly increased amylases activities, while activities of proteases, catalases and peroxidases were, generally, decreased.

In lupin plants, treatment with Cd increased catalytic activity, but activities of amylases, proteases and peroxidases were, in most cases, insignificantly affected. Under the application of Pb, catalytic activity was increased; proteolytic activity was decreased while

activities of amylases and peroxidases were, mostly, insignificantly affected.

Concerning the effect of both Cd and Pb on enzyme activity, Kumar and Banerji (1992) found that, in *Cicer arietinum*, activities of peroxidases and proteases were decreased while amylase activity was increased in response to the application of Cd and Pb. They found also that, the activities of all the studied enzymes were decreased in wheat plants grown under the same treatment with Cd or Pb. Malecka *et al.*^[22] reported that, lead causes two types of unfavorable processes in biological system. Firstly, lead inactivates several enzymes by bindings with their SH-groups. Secondly, lead ions, similarly as those of other heavy metals, can intensify the processes of reactive oxygen species production leading to oxidative stress. Przymusinski *et al.*^[26] reported that, activity of peroxidases in every tested organ of Pb-treated lupin plant. Also, Shaaban^[29] working on *Phaseolus vulgaris* plants, found that, treatment with Cd (25 μM) increased the activities of catalase, superoxidase dismutase and ascorbate peroxidase enzymes. However, Kasim^[15] found that, Cd and Cu induced reduction in the activities of catalases, peroxidases in 25-day-old broad bean plants.

It has been found that, in the present investigation that the combined treatment of (Cd + GA₃) caused, in broad bean plants, significant increase in the amylolytic activity, while activities of proteases, catalases and peroxidases were significantly decreased. In lupin plant,

Table 4: Effect of Cd, Pb, GA₃ and their interactions on the enzyme activity of broad bean and lupin plants. Each value is mean of 3 replicates ± standard error of means.

Broad bean plants				
Treatments	Amylase (mg/ g. F.wt)	Protease (mg/ g. F.wt)	Catalase (µg/ g. F.wt)	Peroxidase (mg/ g. F.wt)
Control	0.12 ± 0.01	2.20 ± 0.33	0.17 ± 0.03	0.65 ± 0.03
Cd	0.27 ± 0.02 **	2.41 ± 0.12	0.48 ± 0.05 **	1.33 ± 0.04 **
Pb	0.29 ± 0.02 **	0.47 ± 0.02 **	0.04 ± 0.02 *	0.12 ± 0.07 **
GA ₃	0.11 ± 0.02	1.14 ± 0.20	0.58 ± 0.03 **	0.19 ± 0.03 **
Cd + GA ₃	0.67 ± 0.05 **	0.42 ± 0.01 **	0.03 ± 0.01 *	0.03 ± 0.02 **
Pb + GA ₃	0.42 ± 0.03 **	2.37 ± 0.11	0.51 ± 0.06 **	1.79 ± 0.07 **
Lupin plants				
Treatments	Amylase (mg/ g. F.wt)	Protease (mg/ g. F.wt)	Catalase (µg/ g. F.wt)	Peroxidase (mg/ g. F.wt)
Control	0.07 ± 0.02	0.75 ± 0.02	0.15 ± 0.04	1.44 ± 0.43
Cd	0.10 ± 0.03	0.67 ± 0.15	0.31 ± 0.01 *	1.42 ± 0.55
Pb	0.08 ± 0.02	0.34 ± 0.02**	0.34 ± 0.04 *	1.44 ± 0.17
GA ₃	0.36 ± 0.02 **	0.97 ± 0.10	0.30 ± 0.02 *	3.61 ± 0.38 *
Cd + GA ₃	0.44 ± 0.02 **	0.45 ± 0.11	0.70 ± 0.06 **	1.52 ± 0.68
Pb + GA ₃	0.21 ± 0.03 *	0.32 ± 0.08**	0.42 ± 0.03 **	1.11 ± 0.33

*Significant at 5% confidence level; **Significant at 1% confidence level.

the same treatment (Cd + GA₃) significantly increased activities of amylases and catalases while, activities of proteases and peroxidases were, generally, insignificantly affected.

The obtained results revealed also that, treatment with (Pb + GA₃) caused in broad bean plants, significant increases as regards the activities of amylases, catalases and peroxidases while, activities of proteases were insignificantly affected especially at the second stage of growth. In lupin plants, treatment with (Pb + GA₃) caused significant increases in the activities of amylases and catalases, significant decrease in proteolytic activity and insignificant changes in the activities of peroxidases. It could be deduced that, the increases observed as regards amylases activities of the two tested plants in response to the combined treatments of GA₃ with either Cd or Pb may be related to the lower levels of carbohydrates resulted by the same treatments. Also, plants that have higher levels of proteins revealed lower activity of proteases. This indicates that, close relationships may be found between the levels of both carbohydrates and proteins on one side and activities of either amylases or proteases on the other side.

REFERENCES

1. Abd El-Hamid, M.F., Nabrawy, A. Amera A.A. Gaber and M. Kamil, Amal, 2003. Control the physiological stress resulted from Cd and Pb foliage application on sugar beet plant by using foliar spray with certain growth regulators. J. Agric. Sci., Mansoura Unvi., 28(5): 3571-3602.
2. Abdel-Hameid, M.E., 2004. Physiological studies on some plants. M.Sc. Thesis Fac. Sci., Al-Azhar Univ.
3. Afifi, W.M., A. Zeinab and Abd M.F. El-Hameid, 1986. Effect of gamma-irradiation on pea seedlings. Ann. Agric., Moshtohor, 24(4): 2047-2057.
4. Ali, E.F., 2006. Influence of zinc and cadmium on the growth and biochemical compounds of wheat plant grown in sand culture. M.Sc. Thesis, Fac. Agric. Alexandria Univ.
5. Bergmeyer, H.U., 1974: Methods of Enzymatic Analysis 1. Seacond es. Academic press. New York.
6. Bialecka, B. and J. Kepezynski, 2007. Changes in concentrations of soluble carbohydrates during germination of *Amaranthus caudatus* L. seeds in relation to ethylene, gibberellin GA₃ and methyl jasmonate. Plant-Growth-Regulation, 51(1): 21-31.
7. Chen, Y., X.X.D. Cao, Y. Lu, and X.R. Wang, 2000. Effects of rare earth metal ions and their EDTA complex on antioxidant enzymes of fish liver Bull. Environ. Contam. Toxicol., 65: 357-365.
8. Eleiwa, M.M.E., 2004. Response of lupin and wheat to soil contamination with heavy metals. Egyptian Journal of soil science, 44(1): 1-17.

9. El-Fouly, M.M., R. Saker, M.D. Fouda, A.M Zaher and A.F. Fawzi, 1988. Effect of GA₃, CCC and B₉ on morphophysiological characters and yield of kidney beans (*Phaseolus vulgaris*). J. Agron. Crop. Sci., 160(2): 94-101.
10. El-Khateeb, A.A., 2000. Effect of GA₃ and IAA sprays on vegetative growth, total yield and photosynthetic pigments in strawberry (*Fragaria ananassa*) J. Agric. Sci. Mansoura Univ., 25(3): 1767-1771.
11. El-Safie, S.F., 2000. Effect of organic matter addition on growth and nutrients uptake by some plants grown on Pb-polluted alluvial soils. Minufia J. Agric. Res., 256(1): 203-215.
12. Ewais, E.A., 2004. Regulation of cadmium, cobalt, nickel and lead accumulation by plant growth regulators in *Ricinus communis* L. cotyledon cultured in vitro. Egypt. J. Biotechnol., 16: 228-240.
13. Gaber, A.M., O.A. El-Shahaby, A.A. Ramadan, 2000. Effect of some hormonal treatments on chemical composition and fauvisism causative agents in the yielded seeds of *Vicia faba*. Egyptian-Journal of Physiological Sciences, 24(1): 17-45.
14. Guo, D.S., Y.Y. Xi, A.Y. Wang, J. Zhang and X.Y. Yuan, 1999. Contribution of an auxin to the uptake of nickel and cadmium in maize seedlings. Biochemical and Environmental science, 12(3): 170-176.
15. Kasim, W.A., 2005. The correlation between physiological and structural alteration induced by copper and cadmium stress in broad bean (*Vicia faba* L.). Egyptian J. of Biol., 7: 20-32.
16. Kevresan, S., N. Petrovic, M. Popovic and J. Kanolrac, 2001. Nitrogen and protein metabolism in young pea plants as affected by different concentrations of nickel, cadmium and lead and molybdenum. J. Plant Nutrition, 24(10): 1633-1644.
17. Khalaf, S.M., Khafagi, A. Om-Mohamed and El-Lawendy, I. Waffa, 1986. Effects of GA₃ and CCC on germination and growth of soybean, common bean, cowpea and pigeon pea plants grown under different levels of salinity. Annals of Agric. Sci. Moshtohor, 24(4): 1965-1982.
18. Kumar, S. and D. Banerji, 1992: Effect of some heavy metals on (in vitro) activities of certain enzymes. Plant Physiology and Biochemistry, New Delhi, 19(1): 33-35.
19. Lindim, C., M.O. Varennes and A.M. Mota, 2001. Remediation of sandy soil artificially contaminated with cadmium using a polyacrylate polymer, Commun. Soil Sci. Plant Anal., 32(9&10): 1567-1574.
20. Lowery, O.H., N.J. Rosebrough, A.L. Farr and R.J. Randall, 1951. Protein measurement with the folin reagent. J. Biol. Chem., 193: 265-275.
21. Marghany, H.A., 2005. Comparative studies on the effect of *Adiantum capillus-veneris* and auxin application on growth of carrot plants cultivated under lead polluted soil. M. Sc. Fac. Sci. Al-Azhar Univ. Cairo. Egypt.
22. Michalska, M. and H. Asp, 2001. Influence of Pb and Cd on growth, heavy metal uptake and nutrient concentration of three lettuce cultivars grown in hydroponic culture. Soil Sci., Plant Anal., 32(3&4): 571-583.
23. Neelue, R.A., Y.B. Gao, S. Liu and A.Z. Ren, 2000. Effect of Cr, Cd and Pb on free praline content in leaves of *Brassica chinensis* L. Chinese. J. of applied and Enviro. Biolo., 6(2): 112-116.
24. Ong, P.S. and G.M. Gaucher, 1972. Protease production by thermophilic fungi. Can. J. Microbiology, 19: 129-133.
25. Picazo, I., R. Ros and J.L. Moya, 2007. Heavy metal hormone interactions in rice plants: Effect on growth, net photosynthesis and carbohydrate distribution. J. Plant growth Reg., 14: 61-67.
26. Przymusinski, R., R. Sobkowiak, B. Ilska and A. Edward, 2007. Organospecific responses of lupin seedling to lead localization of hydrogen peroxide and peroxidase activity. Acta Physiol. Plant, 29: 411-416.
27. Saffan, Samia E., 2008. Alleviation of heavy metal toxicity on *Lupinus termis* plants by kinetin, bentonite and ascorbic acid. Egypt. J. Biotechnol., 28: 135-147.
28. Sayed, A.S., 1999. Effect of lead and Kinetin on growth and physiological components of safflower, Plant Growth Regulation, 29: 167-174.
29. Shaaban, Lamis D., 2008. Interactions between cadmium and zinc in *Phaseolus vulgaris* L. sp. *Nibraska*. Egypt. J. Biotech., 28: 198-209.
30. Snedecor, G.M. and W.G. Cochran, 1982. Statistical methods 7th edition, Iowa state Univ., Press, Ames., Iowa U.S.A., pp. 325-330.
31. Streit, B. and W. Stumm, 1993. In plants as biomonitors. indicators for heavy metals in the terrestrial environment. Markert, B. (Ed.) Weinheim: VCH; pp: 167-178.
32. Umbriet, W.W., R.H. Burris, J.F. stauffer, P.P. Cohen, W.J. Johsen, Lee G.A. page, V.R. patter and W.C. Schneicter, 1969 Manometric techniques, manual describing methods applicable to the studs of tissue metabolism. Burgess publishing Co., U.S.A; pp: 239.
33. Vernon, L.P. and G.R. Selly, 1966. The chlorophylls. Academic press. New York and London.