

Bacteriological Study on Dental Caries and Test their Sensitivity to Virulence Factors and Heavy Metals

^{1,2}Muna Jalal Ali, ^{1,3}Essam A. Makky and ¹Mashitah M. Yusoff

¹Faculty of Industrial Sciences & Technology, Universiti Malaysia Pahang, 26300 Gambang, Kuantan, Pahang, Malaysia

²Department of pathological analyses, Al-Haweeja Technical Institute, Foundation of Technical Education, Kirkuk, Iraq

³Center of Excellence for Advanced Research in Fluid Flow (CARIFF), University Malaysia ,Pahang, 26300 Gambang, Kuantan, Pahang, Malaysia

ARTICLE INFO

Article history:

Received 28 September 2015

Accepted 12 December 2015

Available online 24 December 2015

Keywords:

Bacteria; Heavy metals, Virulence factors. Dental caries

ABSTRACT

Tooth decay is considered the most widespread infectious disease in the world. This study aims to isolate and identify the important bacteria related to tooth decay, determine the sensitivity of bacteria in certain types of antimicrobial agents, and study the effect of heavy metals on bacterial isolates. A total of 50 swabs were collected from the mouths of patients from both gender, with ages ranging from 3–60 years. The patients were advised to consult with dental clinics and specialized centers to isolate and identify the causative agents associated with oral diseases. Results showed that infection rates in younger age groups (3–20 and 20–40) are higher than the elder group (40–60), with percent incidence of 44% and 32%, respectively. Heavy metals sensitivity test against the oral isolates were found lower effect for isolates against (15) heavy metals, where it showed resistance to Iron 3.38% , then nickel, aluminum ,copper, lead to 20.33%,22.03%,27.11%,28.81%respectively, also silver shown 57.62%. And, this similarity were found have sensitive to antimony and chromium 61.01%.while appeared sensitivity to mercury and cadmium by 100, 86.44%. Hemolysin had the highest ability to produce virulence factors (72.88%), followed by lecithinase (42.37%) and protease (25.42%). Lipase and urease had the lowest virulence factor production (10.16%).

© 2015 AENSI Publisher All rights reserved.

To Cite This Article: Muna Jalal Ali, Essam A. Makky and Mashitah M. Yusoff., Bacteriological Study on Dental Caries and Test their Sensitivity to Virulence Factors and Heavy Metals. *Adv. Environ. Biol.*, 9(27), 301-306, 2015

INTRODUCTION

Tooth decay is one of the most common infectious diseases affecting millions of people globally [26]. One of the occasional factors for the disease is dental biofilm, which is the bacterial charge that forms permanently on the tooth surfaces [15]. Hazard factors include unsuitable salivary flow, low quality of salivary buffer, incomplete fluoride exposure, and increased consumption of sugar [14]. Caries indicates the centralized removal of susceptible dental hard tissues by acidic products from the bacterial fermentation of dietary carbohydrates [18]. Tooth decay is a chronic disease that is slowly developing in people. Tooth decay presents as smooth holes and fissured surfaces on the crown and root of a tooth. According to the World Health Organization, 60–90% of school children worldwide have dental cavities [15]. This decay is the result of the interaction of the oral microflora plaque, the tooth surface, nourishment, and the oral environment over time, causing destruction of the tooth enamel [13]. Recently, disease incidence for cavities is decreasing in industrialized nations but is increasing in developing nations [10]. The spread of caries is uneven across the population and communities. The highest incidence is in the lower socioeconomic groups, having limited access to adequate oral health care [8]. Despite the decline in incidence of caries, the United States of America is spending 10 billion USD each year on tooth decay treatment [7]. In other industrialized nations, such as the United Kingdom and China, caries prevalence in the past has been over 50% in children. In developing countries, where oral health care is low, caries are increasing in an alarming rate. Previous studies done in Peru, Mexico, the Philippines, and Taiwan found caries in 75–90% of children [4]. Mutants Streptococci, a group of cariogenic bacteria, is associated in the initiation of dental caries [2]. Another group of bacteria that is substantial in the development of caries is Lactobacillus. Lactobacillus does not usually colonize the tooth surface, but is commonly found in the oral

Corresponding Author: Essam A. Makky, Faculty of Industrial Sciences & Technology, Universiti Malaysia Pahang, 26300 Gambang, Kuantan, Pahang, Malaysia
Tel: +60139553169; Fax: +609-5492766; E-mail: essam22001@gmail.com

cavity including the dorsum of the tongue [26]. Although it could have a significant role in the caries advancement, *Lactobacillus* is not essential in the initiation of dental caries [21]. Positive association between salivary levels and bacterial caries is relevant to carbohydrate exhaustion. The presence of *Streptococcus* and *Lactobacillus* may potentially indicate the occurrence of not only caries but also of carbohydrate consumption [2]. *Streptococcus mutans* is commonly accepted as one of the most substantial etiologic agents in caries development and has been shown to directly cause caries in germ-free and specific pathogen-free rat models. However, the presence of caries has been found even in the absence of *S. mutans*. Although a high percentage of *S. mutans* has been recovered from teeth without caries, *S. mutans* remains the species that is most associated with caries. In gnotobiotic and specific germ-free rodent models, *S. mutans* has the potential to generate caries. Despite the various properties in *S. mutans* that raises its cariogenicity, strong biofilm indicating the presence of dietary sucrose is a stringent component in the development of caries.

Thus, this study aims to isolate and partially identify important bacteria related to tooth decay and diseases of the mouth, determine the susceptibility of bacteria to certain types of antimicrobial agents, study the effect of some heavy metals for bacterial isolates, and study the ability of bacterial isolates in producing some of the virulence factors.

MATERIALS AND METHODS

Isolation of microbial isolates from patients:

Collection of samples:

With the assistance of dentists, specimens in this study have been collected from the dental units in health centers and dental clinics in Gambang, Pahang, Malaysia. Sterile swabs were used for the patients of both genders, with ages ranging from 3–60 years. Collected samples were transferred to the laboratory of Universiti Malaysia Pahang.

Microbial culture:

Samples from the mouth of patients were cultured on nutrient agar plates and were incubated at 37° for 24 hour. The samples were then purified and cultured on agar slants. These were kept in the chiller until use.

Antimicrobial activity test using disc diffusion method:

Heavy metals activity test:

Prepare concentration: Prepared concentration was prepared by using 10 milligram /liter for the ten heavy metals (i.e., Aluminum, Antimony, Cadmium, Chromium, Copper, Copper sulfate, Iron, Iron chloride, Lead, Zinc chloride, Nickel, Silver nitrate, Lead acetate, Mercury, Silver) the stock solution was prepared for concentration. Filter paper disc was used and laden with 25 µl of heavy metal. According to the manufacturer's recommendations, the Muller Hinton agar were prepared and were autoclaved at 121°C for 15 min. The medium was then cooled to 45–50 °C and poured onto the plates. The heavy metals discs were allowed to set on a level surface to a depth of approximately 4 mm. Inoculums from primary culture plates were prepared by touching 3–5 colonies with a swab and transferring them into a plate. The inoculums were mixed with two drops of sterile distilled water and were spread in three plates. The ten heavy metals discs prepared were placed onto the inoculated plates. Subsequently, they were placed in the chiller for 15 min and were incubated at 37 °C. After an overnight incubation, the diameter of each inhibition zone was measured and recorded in mm .

Virulence factors:

Haemolysin:

Hemolysin test was used to investigate the production of blood enzyme. The hemolytic activity of bacteria was assayed by using nutrient agar containing 5% blood. Bacterial isolate cultures were incubated at 37 °C for 24 h on blood agar plates. The appearance of a transparent zone around the bacteria indicates a positive result for hemolysin [21].

Protease:

Skim milk agar medium was used to investigate the production of protease enzyme. The medium was prepared by mixing 100 ml of nutrient agar and 1 ml of sterile skim milk. The mixture was autoclaved to make it sterile and then poured into sterile dishes (Stukus, 1997) Inoculums from primary culture plates were prepared by brushing 3–5 colonies via loop and transferring them onto the plates. The inoculums were incubated for 24 h at 37 °C. Decomposition on areas was observed

Lipase and Lecithinase:

Egg yolk agar was prepared by mixing 100 ml of nutrient agar, which was sterilized via autoclave and was left to cool to 45 °C, with 5 ml of egg yolk. The agar was poured into sterile dishes. The agar was used to distinguish the bacteria that produce lipase or lecithinase enzyme (Cruickshank *et al.*, 1975). Egg yolk agar was

inoculated with colonies of pure isolated bacteria and was incubated at 37 °C for 24–48 h. Egg yolk agar is inferred to be effective on inhibiting lecithinase enzyme around the developing colonies. Egg yolk agar is also used to detect the effectiveness of lipase enzyme. Egg yolk agar test was conducted by immersing the dish in sufficient quantity of a saturated copper sulfate for 20 min. After the removal of excess solution, the dish was dried using the incubator for 30 min. Decomposition of fat by lipase enzyme was indicated by the emergence of greenish blue color in growth areas.

Urease Test:

This test was done to investigate the ability of bacteria to produce urease enzyme and to analyze the urea of ammonia and carbon dioxide content. Urea agar was inoculated and incubated at 37 °C for 18–24 h. Positive result was considered to be indicated by the change in color of the media to pink [9].

RESULTS AND DISCUSSION

Patient's isolates:

Data on bacterial and yeast (59) isolates during the primary isolation of samples are shown in table 1 and figures 1. Data were obtained from the mouths of 50 patients of different ages and genders, composed of 54% males and 46% females. The 20–40 and 3–20 age group were the more infected, with 44% incidence, compared with the elder age group (40–60 years), with 32% incidence. This study confirmed that children and younger individuals are more susceptible to mouth infection compared with other age groups. This finding may be due to the low immunity and low health consciousness of these age groups, as well as due to other factors related to nutrition and public health that increases the rates of infection among them. In another study, [17] stated that children are more susceptible to decay causing bacteria than other age groups are. Infected children who have malformed teeth showed high mortality rates. The frequent sugar consumption of children plays an important role in infections. Mothers can also transfer diseases from their infected teeth to their children. In such case, the levels of bacteria found at the children are similar with that of the mothers.

Table 1: Primary isolation of samples and percentages.

| Patients Samples & age (year) | Isolate number | Percentage (%) |
|-------------------------------|----------------|----------------|
| Single isolate | 33 | 55.93 |
| Mixed isolate | 26 | 44.07 |
| 3-20 | 16 | 32 |
| 20-40 | 22 | 44 |
| 40-60 | 12 | 24 |

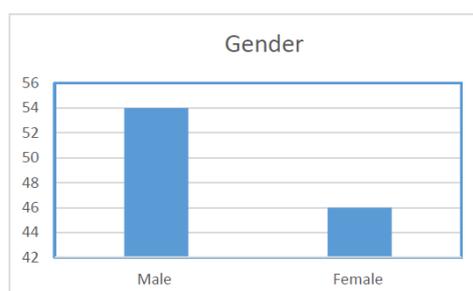


Fig. 1: Percentage of isolates according to gender.

Sensitivity of bacteria to heavy metals:

Sensitivity of bacteria to heavy metal .The results as shown in Figure. 2 that the resistance and sensitive percentages of bacterial isolates fifteen heavy metals, where it showed explain the resistance to heavy metals, Iron 3.38% , then nickel , aluminum ,copper, lead to 20.33%,22.03%, 27.11%, 28.81% respectively, also silver shown 57.62%. And, this similarity were found have sensitive to antimony and chromium 61.01%.while appeared sensitivity to mercury and cadmium by 100%, 86.44%

The results of this study also showed the high resistance shown by the bacterial isolates to iron. The immobilized bacteria performed well iron reduction than free bacteria even under unfavorable pH and temperatures. Evidenced by the results of the current study, the majority of bacterial isolates possessed prescription relatively high sensitive to mercury. Mercury is additionally the sole microorganism metal

resistance system whose mechanism ends up in large scale transformation of its target. The mechanisms of different ion to resistances are supported effluent pumps or living thing sequestration. Barkay and Miller (2003) studied bacterial mercury resistance from atoms to ecosystems so reported that one or more proteins apparently. [5]. The data obtained during this study clearly shows that with sensitive microorganism of cadmium. This may be due largely to reasons related to less concentration from cadmium increase the rates of sensitive isolates

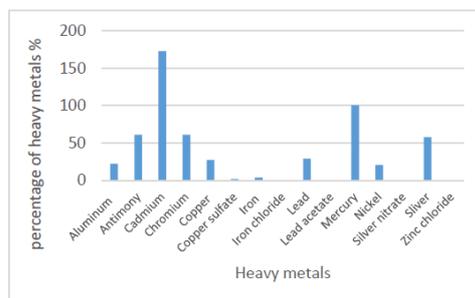


Fig. 2: Percentage of bacterial isolates sensitive of heavy metals.

Virulence Factors:

Figure 3 shows the percentage of bacterial isolates produced to five virulence factors. Hemolysin had the highest production to virulence factors with 72.88%, followed by lecithinase and protease with 42.37%, and 25.42% respectively. Less bacterial isolates were produced to virulence by lipase and urease (10.16%).

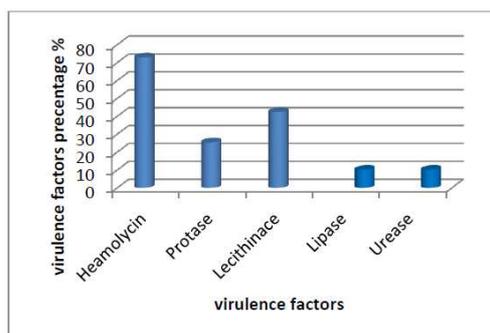


Fig. 3: Percentage to bacterial isolate produced virulence factors.

Virulence is the degree of pathogenicity exhibited by most pathogens and is a measure that effectively differentiates pathogenic and nonpathogenic strains. The degree of virulence depends on several virulence factors. In this study, the most significant result was that of hemolysin at 72.88%. A direct relationship between bacterial isolates and hemolysin was not observed. Bacterial isolate strains that are Gram-positive are noted to contain the highest number of Gram-positive bacteria with much hemolysin produced. Other authors have also shown that 89% of hemolysin produces clinical isolated strains [3]. Takahashi *et al.* [21] showed that 80% of produced hemolysin from the human body is positive of *Aeromonas trota* [21]. Almost 95% of isolated human *Streptococcus* produces a characteristic hemolysin that is only among *Streptococci*. Meanwhile, the second highest virulence factor produced in bacterial isolates was lecithinase at 42.37%. The phospholipid lecithin is one of the chief components of the cell membrane, which can be degraded by lecithinase enzyme, thus producing diglyceride and phosphorylcholine and causing toxicity. Sharaf *et al.* [19] reported that 53 isolates from 60 bacterial isolates were positive of lecithinase when lecithinase-producing bacteria from commercial and homemade foods were studied. [19] Bacterial proteases are recognized as virulence factors in a number of infectious diseases due to their cell and tissue damaging effects. In one study, in which the protease result was 25.42%, a connection was found between the increase in protease production by *Staphylococcus epidermidis* and the obscurity of *Staphylococcus aureus* in biofilms obtained from the same patient [25]. Batra and Walia [5] reported that 39 strains of bacteria-producing protease out of 57 strains were isolated from different soil samples from a cotton field [6]. The lowest percentage of virulence factors in the current study was recorded at 10.16% for both urease and lipase. Urease activity was negatively associated with sugar consumption. In addition, urease activity in saliva increased with age and was positively associated with the levels of *S. mutans* in saliva and with the educational level of the parents. Lipase is a triacylglycerol hydrolyzing enzyme that catalyzes the hydrolysis of water-insoluble free fatty acid and glycerols. Lipase also has a wide range of chemical reactions.

The results of this study are similar to those of Thomas *et al.*, in which they found that *Bacillus mycoides* showed a growth or production of lipase at temperatures below 10 °C or above 50 °C [24]. Joseph [12] reported that sodium chloride increased lipase production, whereas the presence of metals in the media had an inhibitory effect. *S. epidermidis* immobilized cells in agar beads and increased lipase production by 3% compared with free cells [12].

Results of the study showed that the rate of tooth caries was highest in the second age group 44%. The results of this study showed an increase in the proportion of resistance all heavy metals except mercury (100%), cadmium (86.44%) and copper sulfate (1.69%). The highest ability to produce virulence factors was hemolysin 72.88%, lecithinase 42.37 and protease 25.42%, lipase and urease were 10.16%.

Conclusion:

In this study the higher dental caries infection was in younger age stage groups, five of heavy metals were resistance bacterial isolates and hemolysin had the highest ability to produce virulence factors.

ACKNOWLEDGEMENT

The authors gratefully acknowledge University Malaysia Pahang (UMP), Malaysia for the financial supported by grant GRS 140318 that enables the authors to accomplish this work.

REFERENCES

- [1] Ali, M., E.M. Makky, T. Batool, M. Yusoff, 2015. Susceptibility of oral bacteria to antimicrobial agents and virulence factors. *Journal of Chemical & Pharmaceutical Research*, 7(3): 1822-1829.
- [2] Ali, M., E.A. Makky, M.M. Yusoff, 2015. Oral bacteria: Antimicrobial and virulence. *Journal of Chemical & Pharmaceutical Research*, 7(3): 1816-1821.
- [3] Anacarso, I., C. Condò, C. Sabia, P. Messi, S. Niederhausern, M. Bondi and R. Iseppi, 2013. Antimicrobial Resistance and Other Related Virulence Factors in *Staphylococcus Spp* isolated from Food. *Environmental and Humans in Italy. Universal Journal of Microbiology Research*, 1(1): 1-9.
- [4] Bagramian, R.A., F.Garcia-Godoy, A.R. Volpe, 2009. The global increase in dental caries. A pending public health crisis. *Am J Dent*, 22(1): 3-8.
- [5] Barkay, T., S.M. Miller and A.O. Summers, 2003. Bacterial mercury resistance from atoms to ecosystems. *FEMS microbiology reviews*, 27(2-3): 355-384.
- [6] Batra, N., M. Walia, 2014. Production and characterization of alkaline protease from bacteria strains isolated from cotton field. *African Journal of Microbiology Research*, 8(7): 702-709.
- [7] Benjamin, R.M., 2010. Oral health: the silent epidemic. *Public health reports*, 125(2): 158.
- [8] Bowen, W.H., 2002. Do we need to be concerned about dental caries in the coming millennium? *Critical Reviews in Oral Biology & Medicine*, 13(2): 126-131.
- [9] Brown, A.E., 2009. *Benson's Microbiological Applications: Laboratory Manual in General Microbiology, Short Version 11th ed.* USA: McGraw-Hill Higher Education.
- [10] Chu, C., E. Lo, 2008. Promoting caries arrest in children with silver diamine fluoride: a review. *Oral health & preventive dentistry*, 6(4).
- [11] Cruickshank, R., J.P. Duguid, B.P. Marmion, R.H. and Swain, 1975. *The Practical of medical microbiology 12th ed.*: Churchill livingstone.
- [12] Joseph, B., P.W. Ramteke, P.A. Kumar, 2006. Studies on the enhanced production of extracellular lipase by *Staphylococcus epidermidis*. *Journal of General and Applied Microbiology*, 52(6): 315-320.
- [13] Lynch, D.J., 2010. An analysis of the role of glucan-binding proteins in *Streptococcus mutans* biofilm architecture and caries development. (Ph.D.), University of Iowa, USA.
- [14] MejÅre, I., S. Axelsson, G. Dahlén, I. Espelid, A. Norlund, S. Tranæus, S. Twetman, 2014. Caries risk assessment. A systematic review. *Acta Odontologica Scandinavica*, 1-11.
- [15] Petersen, P.E., 2008. World Health Organization global policy for improvement of oral health-World Health Assembly 2007. *International dental journal*, 58(3): 115-121.
- [16] Petersen, P.E., D. Bourgeois, H. Ogawa, S. Estupinan-Day, C. Ndiaye, 2005. The global burden of oral diseases and risks to oral health. *Bulletin of the World Health Organization*, 83(9): 661-669.
- [17] Rao, G.G., 1998. Risk factors for the spread of antibiotic-resistant bacteria. *Drugs*, 55(3): 323-330.
- [18] Selwitz, R.H., A.I. Ismail, and N.B. Pitts, 2007. Dental caries. *The Lancet*, 369(9555): 51-59.
- [19] Sharaf, E.F., W.S. El-Sayed, R. Abosaif, 2014. Lecithinase-Producing Bacteria from Commercial and Homemade Foods: Evaluation of Toxigenic Properties and Identification of Potent Producers. *Journal of Taibah University for Science*, 8(3): 207-215.
- [20] Stukus, P.E., 1997. *Investigating microbiology: a laboratory manual for general microbiology*: Saunders College Publishing.

- [21] Takahashi, E., H. Ozaki, Y. Fujii, H. Kobayashi, H. Yamanaka, S. Arimoto, K. Okamoto, 2014. Properties of Hemolysin and Protease Produced by *Aeromonas trota*. *PloS one*, 9(3): e91149.
- [22] Takahashi, N., B. Nyvad, 2008. Caries ecology revisited: microbial dynamics and the caries process. *Caries research*, 42(6): 409-418.
- [23] Takahashi, N., B. Nyvad, 2011. The Role of Bacteria in the Caries Process Ecological Perspectives. *Journal of Dental Research*, 90(3): 294-303.
- [24] Thomas, A., M. Mathew, A. Valsa, S. Mohan and R. Manjula, 2003. Optimisation of growth conditions for the production of extracellular lipase by *Bacillus mycoides*. *Indian Journal of Microbiology*, 43(1): 67-69.
- [25] Vandecandelaere, I., P. Depuydt, H.J. Nelis and T. Coenye, 2014. Protease production by *Staphylococcus epidermidis* and its effect on *Staphylococcus aureus* biofilms. *Pathogens and disease*, 70(3): 321-331.
- [26] Wongkamhaeng, K., O. Poachanukoon, and S. Koontongkaew, 2014. Dental caries, cariogenic microorganisms and salivary properties of allergic rhinitis children. *International Journal of Pediatric Otorhinolaryngology*. 78(5): 860-865.